



BIOLOGY AND LIFETABLE OF DIAMOND BACK MOTH *PLUTELLA XYLOSTELLA* (L) ON CRUCIFEROUS CROPS

SMRITI SHARMA¹, ANUBHA DAS², JYOTSNA SINGH³, PAVITRA SHARMA⁴, PRIYA SINGH⁵ AND SANJIV MULLICK^{6*}

¹Department of Zoology, Gargi College, University of Delhi, Siri Fort Road, New Delhi 110049, India

²Department of Zoology, Zakir Husain Delhi College (M), University of Delhi,
Jawahar Lal Nehru Marg, New Delhi 110002, India

³Department of Zoology, Daulat Ram College, University of Delhi, Delhi 110007, India

⁴School of Sciences, IGNOU, Maidan Garhi, New Delhi 110068, India

⁵Department of Zoology, Venkateswara College, University of Delhi, New Delhi 110021, India

⁶Department of Zoology, Dyal Singh College, University of Delhi, Lodi Road, New Delhi 110003, India

*Email: sanjivmullick@dsc.du.ac.in (corresponding author): ORCID ID 0000-0002-3156-8868

ABSTRACT

Plutella xylostella L., commonly known as diamondback moth (DBM), is an important pest of cruciferous crops worldwide. In this study, biology and life table parameters of *P. xylostella*, reared on cauliflower (*Brassica oleracea* L. var *botrytis*), cabbage (*B. oleracea* L. var *capitata*) and radish (*Raphanus sativus* L.), were measured to evaluate the suitability of these plants. The developmental period, survivorship and fecundity of *P. xylostella* were statistically same on all three test plants, however longevity of female was significantly higher on cabbage compared to cauliflower and radish. The mortality of larvae was significantly higher during early developmental stages, i.e. first and second larval instars, when reared on cauliflower and cabbage leaves but it was evenly distributed throughout the developmental stages when reared on radish. Moreover, the intrinsic rate of increase (r_m) was highest on cauliflower (0.35) as compared with cabbage (0.29) and radish (0.33), rendering it the most suitable host plant for *P. xylostella*. Present study may be helpful in calculating population dynamics and predicting time of action threshold for the application of insecticides and other control measures for the management of this pest.

Key words: *Plutella xylostella*, biological parameters, cruciferous vegetables, diamondback moth, host suitability, larval period, life table, population dynamics, survivorship.

The diamondback moth (DBM), *Plutella xylostella*, is globally distributed pest, which was reported from India in 1914 (Fletcher, 1914), but now widely distributed in 128 countries (Shelton, 2004). This has been reported to cause 52% yield loss on cabbage in India (Krishnamoorthy, 2004). Globally, this pest has been reported to cause more than 90% of production loss of crucifer crops (Talekar and Shelton, 1993; Furlong et al., 2013; Paudel et al., 2022). The cost for its management was estimated US \$1.0 billion annually in 1993 (Talekar and Shelton, 1993) and US \$5.0 billion in 2012 (Zalucki et al., 2012). The genetic elasticity of *P. xylostella* has enabled it to develop resistance against wide range of insecticides (Sakeel et al., 2017; Uesugi, 2021; Banazeer et al., 2022; Shehzad et al., 2023). This insect has been also reported to develop resistance against biopesticide, *Bacillus thuringiensis* (Ferre and Van Rie, 2002; Sarfraz, 2004). Thus, the management of this pest has become nightmare for farmers (Shelton et al., 1993; Regupathy, 1996; Sarfraz et al., 2006; Furlong

et al., 2013; Philips et al., 2014; Li et al., 2016; Jamtsho et al., 2021; Shehzad et al., 2023).

Life table studies are important conceptual and analytical tools for understanding the population dynamics of insect species (Harcourt, 1969; Rossini et al., 2024). Variation in quality of host-plants affects the growth and development of insects which, in turn, determine their life-history parameters. Even different cultivars and geographical areas of host plants have been observed to affect various life table parameters of *P. xylostella* (Ebrahimi et al., 2009; Hasanshahi et al., 2014; Nazrussalam and Ansari, 2018; Fathipour et al., 2019; Hussain et al., 2019; Jafary-Jahed et al., 2019; Kaur et al., 2021). Thus it is important to know the most vulnerable stage of this pest for its successful management. The biology and life table parameters of *P. xylostella* on various cruciferous crops have been studied in China (Niu et al., 2013), Pakistan (Syed and Abro, 2003; Ramzan et al., 2021) and Iran (Golizadeh et al., 2009; Jafary-Jahed et al., 2019). However,

there is hardly any report about comparative account of various biological and life table parameters of *P. xylostella*, on widely cultivated commercial cultivars of cauliflower, cabbage and radish in India. Present study was undertaken to evaluate and compare suitability of these host plants for *P. xylostella*, and find its most vulnerable stage, for developing cost-effective control strategies.

MATERIALS AND METHODS

The seeds of cauliflower (*Brassica oleracea* L. var. *botrytis*; var. Poosi spl), cabbage (*B. oleracea* L. var. *capitata*; var. Golden acre) and radish (*Raphanus sativus* L.; var. Pusa chetki), were obtained from the National Seeds Corporation, Indian Agriculture Research Institute, New Delhi. Plants were grown from these seeds in the field plots of Zoology Department, University of Delhi, under pesticide free condition, following standard agronomic practices. Leaves from approximately 4-5 weeks old plants at 5-7 leaves stage were used from field plots for the experiments and maintaining culture. *P. xylostella* culture was started with the larvae collected from the cauliflower fields, located on the outskirts of Delhi, and maintained in the laboratory as described by Vats et al., 2019. Insects were reared for 3-4 generations under laboratory conditions for the purpose of acclimatization before using them for bioassay. The culture was replenished at frequent intervals from fresh batch of larvae collected from fields, so as to maintain the vigour of culture. For studying various biological attributes, the freshly laid eggs of *P. xylostella* were taken from culture, sterilized and kept in plastic boxes. After hatching, 10 neonate larvae were transferred with the aid of a fine hair brush on the tender leaves of different test plants, placed in round plastic jars (9 cm diameter x 2 cm height). The bottoms of these jars were lined with moist tissue paper to maintain moisture inside. The larvae were transferred daily into clean plastic jars, provisioned with freshly excised leaves. Observations for survival, mortality and moulting into next instar were recorded daily at a fixed time of the day. Larvae were arranged sex-wise during last instar and kept in separate boxes (7 x 7 cm) for pupation and adult eclosion. The emerging males and females were released in Perspex oviposition cage (20 x 20 x 20 cm) for egg laying. Number of eggs laid by female was recorded daily till all the females stopped laying eggs or died. The experiment was replicated 5 times from different batches of insects, and each replicate consisted responses of 10 larvae.

Stage-specific life table was constructed, following

the methods described by Morris and Miller (1954), to calculate mortality and survival rate at each developmental stage. For calculating the adult survival and reproductive parameters, age of the adults from the time of emergence was taken into account. The age-specific lifetable was constructed based on the method described by Atwal and Bains (1974), and Syed and Abro (2003). Sex-ratios for different host plants were calculated according to the formula, $F/F+M$, where F = number of females and M = number of males. Depending upon the sex-ratio, the numbers of eggs laid by individual female were used to get the number of female birth (mx). The values of pivotal age in days (x), survival of female at age x (lx) and age schedule for female birth at age x (mx), were calculated from the data on life table with respect to each host plant. The number of individuals survived and mean number of female offspring produced at each age interval were recorded. The following life table parameters were calculated from the data: net reproductive rate (R_0) = $\sum lx.mx$, mean generation time (T_c) = $\sum x.lx.mx / R_0$, intrinsic rate of increase (rm) = $\text{Loge}R_0/T_0$ and the finite rate of population increase (λ) = e^{rm} . The various biological and life table parameters were subjected to one way Analysis of Variance (ANOVA) followed by Tukey's post-hoc test. All statistical analyses were performed using the computer software programme, Sigma Stat 2.0. The linear relationship between intrinsic rate of increase (rm) and generation time (T_c) was established by computing R^2 (square of Pearson correlation coefficient) using MS- Excel software program.

RESULTS AND DISCUSSION

There are various reports about influence of host plants on the biological parameters of lepidopteran insect pests (Syed and Abro, 2003; Razmjou et al., 2014; Huaripata and Sanchez, 2019; Kaur et al., 2021; Ramzan et al., 2021; Ahmed et al., 2022; Gopalakrishnan and Kalia, 2022; Mason, 2022; Keerthi and Suroshe, 2024). These findings have been cross-checked in present study on cabbage, cauliflower and radish which indicated significant effects of host plants on the survival, development, longevity and life table parameters of *P. xylostella*. In present study, growth and development of *P. xylostella* varied on different test plants, i.e. cauliflower, cabbage and radish (Table 1). The durations of larval period were 7.8 days on cauliflower, 7.7 days on radish and 8.5 days on cabbage, the difference being not statistically significant ($p > 0.05$). Our observation on larval period is consistent with earlier reports on cabbage (Ahmad et al., 2011; Rasool et al., 2021),

cauliflower (Gowri and Manimegalai, 2017; Jafary-Jahed et al., 2019) and radish (Yang et al., 2021). However, significant differences were observed in the pupal duration for the larvae reared on these test plants. Pupal duration on cauliflower (3.6 days) was significantly lower, as compared to cabbage (4.9 days) and radish (4.9 days) ($p < 0.001$). Our findings are in conformity with earlier reports about cauliflower being the best host plant for *P. xylostella* (Dube and Chand, 1977; Syed and Abro, 2003; Golizadeh et al., 2009). The total number of larvae that successfully developed and survived to emerge as normal adults were statistically equal on all the three host plants.

The quality of host plant species and their cultivars have been reported to affect oviposition of herbivorous insects (Awmack and Leather, 2002). Our study on the ovipositional responses revealed that egg laying by *P. xylostella* females starts from the first day of emergence, which continues till 15th day, the highest being on the second day (Fig. 1). The females lay more than 50% of her eggs in first three days of emergence. The post-ovipositional period (when ≤ 2 eggs per female were laid) lasts for 2-3 days for the females reared during larval stages on cauliflower and radish, whereas those reared on cabbage were significantly longer and extended for 7-8 days. The number of eggs laid by the females,

developed from larvae fed on cauliflower, cabbage and radish leaves were 338.2, 314.8 and 337.1 respectively, the difference being not statistically significant. This indicates equal suitability of all the three cruciferous plants for oviposition of *P. xylostella*. However, Niu et al. (2013) recorded significant differences between fecundity of diamondback moth females reared on cauliflower and radish leaves. We observed significantly higher longevity of females reared on cabbage as compared to those reared on cauliflower and radish (Table 1). The longevity and fecundity of females are considered a 'trade-off' relationship. The increase in fecundity of females has been observed to shorten their longevity (Bell and Koufopanou, 1986; Begum et al., 1996; Thyloor et al., 2016)). Similar significant correlation between adult longevity and fecundity was also observed when *P. xylostella* larvae were reared on radish and pea leaves (Yang et al., 2021). Females from larvae reared on cabbage lived longer than those fed on cauliflower and radish. However, higher longevity of females, developed from larvae fed on cabbage, did not manifest into decrease in oviposition. It appears that these females maintain extra nutrient reservoir to utilize for fecundity, as reported in case of *Spodoptera litura* (Yu et al., 2014).

Shorter developmental time and higher ovipositional potential of an insect on a plant are indicators of its high host suitability (Van Lenteren and Noldus, 1990). Earlier studies on the comparative performance of *P. xylostella* found that radish was not a good host for *P. xylostella* (Dube and Chand, 1977; Singh and Singh, 1982; Saeed et al., 2010). However, present study indicates radish to be equally good host as cauliflower and cabbage, having similar developmental time, survivorship and oviposition. This difference may have been caused due to differences in the cultivars of plant species.

The stage-specific life table constructed in the present study reveals that maximum mortality occurred

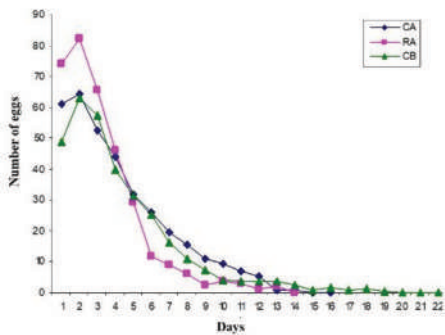


Fig. 1. Ovipositional response of *P. xylostella* in relation to their age. CA, cauliflower; RA, radish; CB, cabbage

Table 1. Biological parameters of *P. xylostella* on three host plants (detached leaf bioassay)

Host plants	Larval duration (days)	Rate of larval pupation (%)	Pupal duration (days)	Rate of survival to adults (%)	Fecundity/ female	Female longevity (days)
Cauliflower	7.83± 0.37 ^a	80± 10.00 ^a	3.61± 0.28 ^a	78± 8.37 ^a	338.17± 26.79 ^a	13.13± 2.67 ^a
Radish	7.66± 1.31 ^a	82± 13.04 ^a	4.96± 0.12 ^b	76± 11.4 ^a	337.11± 9.35 ^a	12.44± 3.63 ^a
Cabbage	8.53± 1.02 ^a	76± 11.40 ^a	4.87± 0.15 ^b	72± 8.37 ^a	314.82± 15.66 ^a	17.61± 4.16 ^b

Values are mean ± SD; Means followed by different lower case letter in a column are significantly different ($p < 0.001$) (one way ANOVA and the means separated by Tukey's test)

during early developmental stages, i.e. first (L1) and second (L2) larval stages, both on cauliflower (16.6%) and cabbage (24.8%). This indicates the vulnerability of this insect during early stages. Thus, control measures applied during early larval stages will be more effective. Mortality was quite low during later developmental stages, i.e. L3, L4 and pupal stage, which accounts 7.26% on cauliflower and 5.26% on cabbage. However, mortality on radish was evenly distributed during the larval and pupal stages (Table 2).

A classical age-specific life table gives a clear understanding of the age dynamics of adult population under controlled laboratory conditions (Carey, 2001). The intrinsic rate of increase (r_m) is an important parameter in life table. This parameter appropriately summarizes the physiological state of insect in

relation to its capacity to increase in number, and thus considered suitable index to evaluate the ability of an insect to colonize on host plants in fields (Southwood and Henderson, 2000). The age-specific life table parameters of *P. xylostella* on different host plants are shown in Table 3. The net reproductive rate of female birth (R_0) was highest on cauliflower (201.1) followed by cabbage (169.2) and radish (162.2), the difference being not significant. The mean generation time (T_c) was significantly higher on cabbage (17.7 days) as compared to cauliflower (15.2 days) and radish (15.3 days) ($p < 0.001$) (Table 3). However, mean generation time on cauliflower and radish was statistically equal ($p > 0.05$). The intrinsic rate of population increase (r_m) was significantly lower on cabbage (0.29) as compared to cauliflower (0.35) and radish (0.33) ($p <$

Table 2. Stage specific life table of *P. xylostella* on three host plants

Stage of the Insect (x)	Host Plant	No. living at the beginning (n_x)	Survivorship (l_x)	No. dying during x (d_x)	Factors responsible for d_x ($d_x f$)	d_x as % of n_x ($100q_x$)	Survival rate within x (S_x)
Eggs	Cauliflower	50	1	0	-	0	1
	Cabbage	50	1	0	-	0	1
	Radish	50	1	0	-	0	1
Larva I	Cauliflower	50	1	6	Inability to come out from the leaf	12	0.88
	Cabbage	50	1	2	Inability to come out from the leaf	4	0.96
	Radish	50	1	1	Inability to come out from the leaf	2	0.98
Larva II	Cauliflower	44	0.88	2	Unknown	4.55	0.95
	Cabbage	48	0.96	10	Unknown	20.8	0.79
	Radish	49	0.98	4	Unknown	8.1	0.92
Larva III	Cauliflower	42	0.84	2	Unknown	4.76	0.95
	Cabbage	38	0.76	0	-	-	1
	Radish	45	0.90	1	Unknown	2.2	0.98
Larva IV	Cauliflower	40	0.80	1	Unknown	2.5	0.98
	Cabbage	38	0.76	0	-	-	1
	Radish	44	0.88	4	Unknown	9.1	0.91
Pupa	Cauliflower	39	0.78	0	-	-	1
	Cabbage	38	0.76	2	Unknown	5.26	0.95
	Radish	40	0.80	2	Pupa Shrunk and dried up	5.0	0.95
Adult	Cauliflower	39	0.78	-	-	-	-
	Cabbage	36	0.72	-	-	-	-
	Radish	38	0.76	-	-	-	-

Table 3. Life table parameters of *P. xylostella* on three host plants

Host plants	R_o	T_c	Rm	$\lambda = e^{rm}$
Cauliflower	201.13± 36.93 ^a	15.22± 0.47 ^a	0.348± 0.006 ^a	1.415± 0.009 ^a
Radish	162.19± 33.33 ^a	15.32± 0.79 ^a	0.332± 0.025 ^a	1.394± 0.034 ^a
Cabbage	169.20± 35.71 ^a	17.70± 0.94 ^b	0.289± 0.021 ^b	1.336± 0.028 ^b

Values are mean ± SD; Means followed by different lower case letter in a column are significantly different; ($p < 0.001$) according to one way ANOVA and the means were separated by Tukey's test.

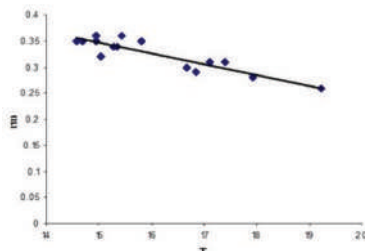


Fig. 2. Intrinsic rate of increase (rm) vs mean generation time (T_c) of *P. xylostella* ($R^2 = 0.906$, $p < 0.001$)

0.001). Similarly, the finite rate of increase (e^{rm}) was significantly lower for cabbage (1.34) as compared to cauliflower (1.42) and radish (1.39) ($p < 0.001$). Multiple linear regression analysis between mean generation time (T_c) (independent variable) and the intrinsic rate of population increase (rm) (dependent variable) yielded $R^2 = 0.906$ ($p < 0.001$) (Fig. 2). Thus, a higher T_c value decreases rm value, indicating a negative effect of T_c on rm. Present study clearly indicates that cauliflower is the most suitable host plant for *P. xylostella* as its intrinsic rate of increase (rm), resulting from faster development (shorter generation time, T_c), higher survivorship and higher oviposition, was highest on this plant. Thus, population growth of *P. xylostella* will be much faster on this crop and will require frequent monitoring for application of effective control measures. Similar finding of high intrinsic rate of increase (rm) of *P. xylostella* on cauliflower was also reported by Saeed et al. (2010). It was also observed that the rm value of *P. xylostella* reared on radish was significantly higher as compared to cabbage. The present study provides a comprehensive account of biological and life table parameters of *P. xylostella* on three brassicaceae crop plants, which will help to understand the performance of this pest in Indian scenario. This information would be useful in developing effective control strategies to keep this pest in check.

ACKNOWLEDGEMENTS

The work was carried out in the laboratory of Prof A K Singh, Department of Zoology, University of Delhi. The authors are thankful to Prof A K Singh for

his supervision and constant guidance for designing experiments and preparation of this manuscript for publication.

AUTHOR CONTRIBUTION STATEMENT

S S contributed to conceptualization and methodology; SS, JS, and P Sh conducted experiments. Analysis of the experimental data was done by S M, A D and S S. S S, A D and P Si wrote the original manuscript. S M reviewed and edited the manuscript. All authors have contributed at different stages of bioassays and approved for the submission of this manuscript for publication.

FINANCIAL SUPPORT

The authors declare that no research grant was involved.

CONFLICT OF INTEREST

No conflict of interest.

REFERENCES

- Ahmad H, Kumar M, Sharma D, Jamwal V V S, Khan R B, and Gupta S. 2011. Bionomics of diamondback moth, *Plutella xylostella* (L.) on cabbage. *Annals of Plant Protection Sciences* 19(1): 80-83.
- Ahmed MA, Cao He-He, Jaleel W, Amir M B, Ali M Y, Smaghe G, Liu Tong-Xian. 2022. Oviposition preference and two-sex life table of *Plutella xylostella* and its association with defensive enzymes in three brassicaceae crops. *Crop Protection* 151.
- Atwal AS, Bains S S. 1974. *Applied animal ecology*. Kalyani publishers, Ludhiana. pp. 245.
- Awmack C S, Leather S R. 2002. Host plant quality and fecundity in herbivorous insects. *Annual Review of Entomology* 47: 814 - 844.
- Banazeer A, Afzal M B S, Hassan S, Ijaz M, Shad S A, Serrão J E. 2022. Status of insecticide resistance in *Plutella xylostella* (Linnaeus) (Lepidoptera: Plutellidae) from 1997 to 2019: cross-resistance, genetics, biological costs, underlying mechanisms, and implications for management. *Phytoparasitica* 50: 465-485.
- Begum S, Ritsuko T, Fujisaki K, Nakasuji F. 1996. The effects of wild cruciferous host plants on morphology, reproductive performance and flight activity in the diamondback moth, *Plutella xylostella* (Lepidoptera: Yponomeutidae). *Research on Population Ecology* 38: 257-263.
- Bell G, Koufopanou V. 1986. The cost of reproduction. In *Oxford Surveys*

- in Evolutionary Biology, ed. R. Dawkins, M. Ridley, 3: 83-131 Oxford, UK: Oxford Univ. Press.
- Carey J R. 2001. Insect biodemography. Annual Review of Entomology 46: 79-110.
- Dube R B, Chand P. 1977. Effect of food plants on the development of *Plutella xylostella* (L.) (Lepidoptera: Plutellidae). Entomon 2(2): 139-140.
- Ebrahimi N, Talebi A A, Fathipour Y. 2009. Demographic parameters of the diamondback moth, *Plutella xylostella* (Lep.: Plutellidae) on five rapeseed cultivars. Journal of Entomological Society of Iran 28: 49-59.
- Fathipour Y, Kianpour R, Bagheri A, Karimzadeh J, Hesseininaveh V. 2019. Bottom-up effects of Brassica genotypes on performance of diamondback moth, *Plutella xylostella* (Lepidoptera: Plutellidae). Crop Protection 115: 135-141.
- Ferre J, Van Rie J. 2002. Biochemistry and genetics of insect resistance to *Bacillus thuringiensis*. Annual Review of Entomology 47: 501-533.
- Furlong M J, Wright D J, Dosdall L M. 2013. Diamondback moth ecology and management: problems, progress, and prospects. Annual Review of Entomology 58: 517-541.
- Golizadeh A, Kamali K, Fathipour Y, Abbasipour H. 2009. Life table of the diamondback moth, *Plutella xylostella* (L.) (Lepidoptera: Plutellidae) on five cultivated brassicaceous host plants. Journal of Agricultural Science and Technology 11: 115-124.
- Gopalakrishnan R, Kalia V K. 2022. Biology and biometric characteristics of *Spodoptera frugiperda* (Lepidoptera: Noctuidae) reared on different host plants with regard to diet. Pest Management Science 78(5): 2043-2051.
- Gowri G, Manimegalai K. 2017. Life table of diamondback moth, *Plutella xylostella* (L.) (Lepidoptera: Plutellidae) on cauliflower (*Brassica oleracea* var *botrytis* L.). Journal of Entomology and Zoology Studies 5(4): 1547-1550.
- Harcourt D G. 1969. The development and use of life tables in the study of natural insect populations, Annual Review of Entomology 14:175-196.
- Hasanshahi G, Jahan F, Abbaasipour H, Salehi-Tabar M, Askarianzadeh A, Karim J, Rahimi A H. 2014. Biology and demography of diamondback moth, *Plutella xylostella* (Lepidoptera: Plutellidae) on five cauliflower cultivars under laboratory conditions. Acta Entomologica Sinica 57(1): 61-66.
- Huaripata C, Sanchez G. 2019. Life cycle of the diamondback moth *Plutella xylostella* L. (Lepidoptera: Plutellidae), in broccoli and cauliflower under laboratory conditions. Peruvian Journal of Agronomy 3(1): 1- 5.
- Hussain R, Mohsin A, Nasir M F, Akram Z, Qureshi M S. 2019. Effect of two cabbage varieties on fitness of *Plutella xylostella* under controlled conditions. University of Sindh Journal of Animal Sciences 3(4): 1-5.
- Jafary-Jahed M, Razmjou J, Nouri-Ganbalani G, Naseri B, Hassanpour M, Leppla N C. 2019. Life table parameters and oviposition preference of *Plutella xylostella* (Lepidoptera: Plutellidae) on six brassicaceous crop plants. Journal of Economic Entomology 112(2): 932-938.
- Jamtsho M, Banu N, Kinley C. 2021. Critical review on past, present and future scope of diamondback management. Plant Archives 21(1): 1199-1210.
- Kaur M, Thakur M, Sharma K C. 2021. Biological and life table parameters of diamondback moth, *Plutella xylostella* (L.) (Lepidoptera: Yponomeutidae) from five different geographical regions of India. Phytoparasitica 49(6): 819-827.
- Keerthi M C, Suroshe S S. 2024. Effect of host plants on the fitness and demographic parameters of the diamondback moth, *Plutella xylostella* (L.) using age-stage, two-sex life tables. Journal of Plant Diseases and Protection 131: 143-154.
- Krishnamoorthy A. 2004. Biological control of diamondback moth, *Plutella xylostella* (Linn.), an Indian scenario with reference to past and future strategies. In: Kirk, A. A., Bordat, D. (Eds). Proceedings of the International Symposium, 21-24 October, 2002, Montpellier, France, CIRAD, 2004, pp. 204-211.
- Li Z Y, Feng X, Liu S S, You M S, Furlong M J. 2016. Biology, ecology, and management of the diamondback moth in China. Annual Review of Entomology 61: 277-296.
- Mason P. 2022. *Plutella xylostella* (diamondback moth), CABI Compendium, 10.1079/cabicompendium.42318
- Morris R F, Miller C A. 1954. The development of life table for the spruce budworm. Canadian Journal of Zoology 32: 283-301.
- Nazrussalam, Ansari M S. 2018. Life table of *Plutella xylostella* (L.) on brassica hosts. Indian Journal of Entomology 80(1): 13-18.
- Niu Y Q, Li X W, Li P, Liu T X. 2013. Effects of different cruciferous crops on fitness of *Plutella xylostella* (Lepidoptera: Plutellidae). Crop Protection. 54: 100-105.
- Paudel A, Yadav P, Karna P. 2022. Diamondback moth *Plutella xylostella* (Linnaeus, 1758) (Lepidoptera: Plutellidae); A real menace to crucifers and its integrated management tactics. Turkish Journal of Agriculture Food Science and Technology 10(12): 2504-2515.
- Philips C R, Fu Z, Kuhar T P, Shelton A M, Cordero R J. 2014. Natural history, ecology and management of diamondback moth (Lepidoptera: Plutellidae) with emphasis on the United States. Journal of Integrated Pest Management 5(3): D1-D11
- Ramzan M, Amin M, Zahid M, Nasir M, bin Umar A. 2021. Effect of different host plants on the biology of diamondback moth, *Plutella xylostella* under laboratory conditions in northern Punjab, Pakistan. Egyptian Academic Journal of Biological Sciences, F. Toxicology and Pest Control 13(1): 45-51.
- Rasool K, Ahmad S B, Wani A R, Yaqoob M, Maqbool S, Rasool K, Mohd. Bhat A. 2021. Seasonal incidence of diamondback moth *Plutella xylostella* (Lepidoptera: Yponomeutidae) on cabbage. The Pharma Innovation Journal 10(11): 793-795.
- Razmjou J, Naseri B, Hemati S A. 2014. Comparative performance of the cotton bollworm, *Helicoverpa armigera* (Hübner) (Lepidoptera: Noctuidae) on various host plants. Journal of Pest Science 87: 29-37.
- Regupathy A. 1996. Insecticide resistance in diamondback moth (DBM), *Plutella xylostella* (L.): status and prospects for its management in India. In Proceedings: The management of diamondback moth and other crucifer pests. pp. 233-239.
- Rossini L, Contarini M, Speranza S, Mermer S, Walton V, Francis F, Garone E. 2024 Life tables in entomology: A discussion on tables' parameters and the importance of raw data. PLoS One 19(3): e0299598.
- Saeed R, Sayyed A H, Shad S A, Zaka S M. 2010. Effect of different host plants on the fitness of diamond-back moth, *Plutella xylostella* (Lepidoptera: Plutellidae). Crop Protection 29(2): 178-182.
- Sakeel M, Farooq M, Nasim W, Akram W, Khan F Z A, Jaled W, Zhu X, Yin H, Li S, Fahad S, Hussain S, Chauhan B S, Jin F. 2017. Environment polluting conventional chemical control compared to an environmentally friendly IPM approach for control of diamondback moth, *Plutella xylostella* (L.) in China: a review. Environment Science Pollution Research. 24: 14537-14550

- Sarfraz M. 2004. Interaction between diamondback moth and *Bacillus thuringiensis*. Outlooks on Pest Management 15: 167-171.
- Sarfraz M, Dosdall L M, Keddle B A. 2006. Diamondback moth–host plant interactions: implications for pest management. Crop Protection 25(7): 625-639.
- Shehzad M, Bodlah I, Siddiqui J A, Bodlah M A, Fareen A G E, Islam W. 2023. Recent insights into pesticide resistance mechanisms in *Plutella xylostella* and possible management strategies. Environmental Science Pollution Research (42): 95296 -95311.
- Shelton A M, Wyman J A, Cushing N L, Apfelbeck K, Dennehy T. 1993. Insecticide resistance of diamondback moth, *Plutella xylostella* (Lepidoptera: Plutellidae) in North America. Journal of Economic Entomology 86: 11-19.
- Singh S P, Singh D. 1982. Influence of cruciferous host plants on the survival and development of *Plutella xylostella* L. Journal of Research (Punjab Agricultural University) 19: 100-104.
- Southwood T R E, Henderson P A. 2000. The construction, description, and analysis of age-specific life-tables. In T. R. E. Southwood & P. A. Henderson (Eds.), Ecological methods. Oxford, UK: Blackwell Science pp. 404-436.
- Syed T S, Abro G H. 2003. Effect of brassica vegetable hosts on biology and life table parameters of *Plutella xylostella* under laboratory conditions. Pakistan Journal of Biological Sciences 6(22): 1891-1896.
- Talekar N S, Shelton A M. 1993. Biology, ecology, and management of the diamondback moth. Annual Review of Entomology 38: 275-301.
- Thyloor R, Kiran T, Doddamane M. 2016. Cost of mating: a study on physiological trade-offs between fecundity and longevity in laboratory reared *Helicoverpa armigera* (Hubner) female moths. Journal of Entomology and Zoology Studies 4(6): 768-771.
- Uesugi R. 2021. Historical changes in the lethal effects of insecticides against the diamondback moth, *Plutella xylostella* (L.). Pest Management Science 77(7): 3116-3125.
- Van Lenteren J C, Noldus L P J J. 1990. Whitefly-plant relationships, behavioral and ecological aspects. In: Gerling, D. (Ed.), Whitefly: their bionomics, pest status and management. Intercept. Andover, England. pp. 47-89.
- Vats T K, Rawal V, Mullick S, Devi M A, Singh P, Singh A K. 2019. Bioactivity of *Ageratum conyzoides* (L.) (Asteraceae) on feeding and oviposition behaviour of diamondback moth *Plutella xylostella* (L.) (Lepidoptera: Plutellidae). International Journal of Tropical Insect Science 39: 311-318.
- Yang F Y, Chen J H, Ruan Q Q, Wang B B, Jiao L, Qiao Q X, He W Y, You M S. 2021. Fitness comparison of *Plutella xylostella* on original and marginal hosts using age-stage, two-sex life tables. Ecology and Evolution 11: 9765-9775.
- Yu J F, Li C, Xu J, Liu H, Ye H. 2014. Male accessory gland secretions modulate female post-mating behavior in the moth, *Spodoptera litura*. Journal of Insect Behaviour 27: 105-116.
- Zalucki M P, Shabbir A, Silva R, Adamson D, Shu-Sheng L, Furlong M J. 2012. Estimating the economic cost of one of the world's major insect pests, *Plutella xylostella* (Lepidoptera: Plutellidae): Just how long is a piece of string? Journal of Economic Entomology 105: 1115-1129.

(Manuscript Received: July, 2024; Revised: October, 2024;
Accepted: October, 2024; Online Published: November, 2024)
Online First in www.entosocindia.org and indianentomology.org Ref. No. e24387